

Cell disruption and extract

Cell disruption is an essential part of biotechnology and the downstream processes related to the manufacturing of biological products.

Several types of cell disruption methods exist, as biological products may be extracellular, intracellular or periplasmic. Cell disruption methods can be categorised into **mechanical** methods and **non-mechanical** methods.

Non-mechanical methods can be divided into physical, chemical and enzymatic methods, Mechanical methods are divided into solid shear and liquid shear methods.

Cell disruption Methods:

A/ Physical methods

1. Disruption in beads mill
2. Disruption using a rotor-stator mill
3. Disruption using French press
4. Disruption using ultrasonic vibrations

B/ Chemicals and phisochemicals methods

1. Disruption using detergents
2. Disruption using enzymes(Lysozyme)
3. Disruption using solvents
4. Disruption using osmotic shock

Factors that influence the selection of disruption method include:

- 1-The susceptibility of the cells to disruption.
- 2-Product stability.
- 3-The ease of extraction from the cell debris.
- 4-The speed of the method and the cost of the method.

- Cell walls act as additional disruption deterrents, with yeast cells being particularly difficult to disrupt, as the cell wall limits the solvents access to the desired products.
- Different cells have different structures; hence they require different methods for disruption.
- Other types of cells requiring disruption are bacterial cells, moulds, plant cells, mammalian cells and ground tissue.
- Mechanical methods produce heat during the process, so additional cooling systems are required when using mechanical cell disruption methods.
- The drying of the cell mass enhances disruption methods and help bring down the costs.

There are many methods of cellular disruption because there are many types of cells.

- 1- Bacterial cells** may have different disruption methods, depending on whether they are gram positive or gram negative, as the amount of peptidoglycan and the presence of an envelope affect the overall process. Bacteria vary by different type of disintegration(digestive enzymes like lysozymes, osmotic shocks, chemical or mechanical treatment.
- 2- Mammalian cells** are the easiest to disrupt as they lack a cell wall.
- 3-Plant cells:** are more difficult to disrupt than animal cells because of the cellulosic walls.

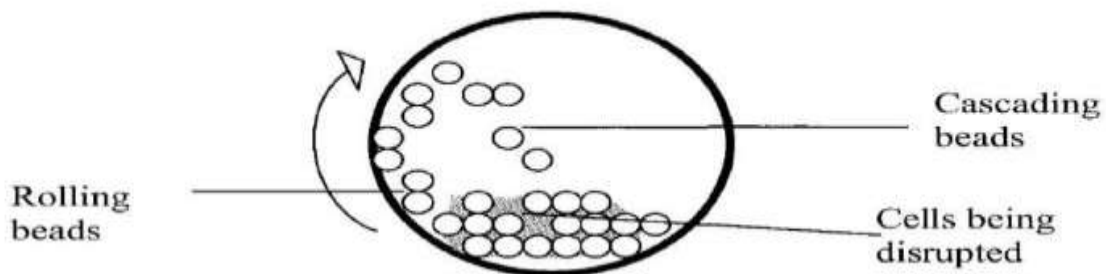
There are three simplest types for disruption of bacterial cells like:

1- Bead mill

All bead devices open the cells or homogenize tissues by throwing the beads against the cells/tissue. Also the accelerated beads generate strong shear in the liquid buffer surrounding the cells/tissues, which also pulls them apart.

Two methods to accelerate the beads by:

- (1) Shaking the entire container
- (2) A spinning agitator within a container



The basic principle of a bead mill

- **Commonly used** for disrupting yeast cells and for grinding animal tissue.
- **The choice of bead size and weight** is greatly dependent on the type of cells.
- **The diameter** can affect the efficiency of cell disruption .
- **An optimal condition for bead load** is considered between 80 and 85% of the free volume. The discs run at a speed of 1500-2250 rpm.

Glass beads with a diameter greater than 0.5 mm are considered best for yeast cells, and diameter lesser than 0.5 mm is optimal for bacterial cells.

The process variables are: agitator speed, proportion of the beads, beads size, cell suspension concentration, cell suspension flow rate, and agitator disc design.

The increased number of beads increases the degree of disruption, due to the increased bead-to-bead interaction. The increased number of beads, however, also affects the heating and power consumption.

1- Disintegration of bacterial cells by glass beads:

1-Suspend bacterial cells in the extraction buffer and mix them with sterilized glass beads then vortex the mixture for 20 min in cold condition.

2- Remove the suspension by Pasteur pipette then centrifuge it at 8000 r/min for 15 min in cooling centrifuge.

3- discard the pellet and collect the supernatant and measure the activity of product in order to do other steps of extraction and purification of our desired product.

Nonmechanical methods: use chemicals to solubilise the components in the cell walls to release the product.

Chemical requirements:

1. Products are insensitive to the used chemicals.
2. The chemicals must be easily separable.

Types of chemicals:

Surfactants (solubilising lipids): sodium sulfonate, sodium dodecylsulfate.

Alkali: sodium hydroxide, harsh.

Organic solvents: penetrating the lipids and swelling the cells. e.g. toluene. e.g. Bacteria were treated with acetone followed by sodium dodecyl sulfate extraction of cellular proteins.

Non mechanical to lyse cell walls to release the product. gentle, but high cost i.e. lysozyme (carbohydrase) to lyse the cell walls of bacteria.

Triton X-100

Triton X-100 is soluble at 25 °C in water, toluene, xylene, ethyl ether, ethyl alcohol, isopropyl alcohol, but insoluble in kerosene.

Triton X-100 is a commonly used **detergent** in laboratories. It is widely used to lyse cells to extract protein or organelles, or to permeabilize the membranes of living cells.

2- TritonX-100 and K_2HPO_4

1- Add (12.5%) of K_2HPO_4 and 2% of Triton x-100 to the suspension of bacterial cells and incubate them at 25°C for 30 min.

2- centrifuge the suspension at 8000 r/min for 30 minute in cooling centrifuge.

3- discard the pellet and collect the supernatant and measure the activity of product in order to do other steps of extraction and purification of our desired product.



Potassium hydrogenphosphate

Potassium hydrogen(tetraoxidophosphate)(2-)

Other names

Potassium monohydrogen phosphate

Phosphoric acid dipotassium salt

Potassium phosphate dibasic



3- Disintegration of bacterial cells by Lysozyme enzyme

1- Suspend bacterial cells in 100ml DW.

2- Add 0.2mg/ml and 200 $\mu\text{g}/\text{ml}$ of lysozyme then incubate them for 20 min. at 37°C.

2- Centrifuged the suspension at 8000 r/min for 15 min in cooling centrifuge.

3- discard the pellet and collect the supernatant and measure the activity of product in order to do other steps of extraction and purification of our desired product.